## Comparative Responses of *Lilium longiflorum*, cv, 'Ace' Pollen to Aflatoxins B<sub>1</sub>, B<sub>2</sub>, and G<sub>1</sub>

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For the past few years we have been examining the possibility that the growth and development of plants and their organs can be used as bioassay materials for the hepatocarcinogenic aflatoxins (DASHEK & LLEWELLYN 1974, 1977; JONES et al. 1980; DANLEY et al. 1981; DASHEK et al. 1979). One of the plant parts that we have utilized rather extensively is Lilium longiflorum, cv. 'Ace' pollen. The reasons for selecting pollen included the ease with which it can be cultured as well as its commercial availability and relatively large size. The simplicity of quantifying the germination and tube elongation responses of in vitro germinating pollen to exogenous compounds is well known.

The four main aflatoxins are AFB<sub>1</sub>, AFB<sub>2</sub>, AFG<sub>1</sub> and AFG<sub>2</sub> (Fig. 1). We have been testing the ability of each of these to affect germination and subsequent tube elongation in order to generate dose-response curves, i.e., log percent germination and/or tube length related to medium toxin concentration.

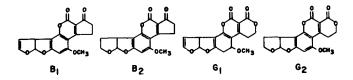


Fig. 1. Structures of AFB<sub>1</sub>, AFB<sub>2</sub>, AFG<sub>1</sub> and AFG<sub>2</sub>.

This paper compares the germination and tube elongation responses of in vitro cultured pollen to AFB, AFB, and AFG,

## EXPERIMENTAL PROCEDURES

Growth conditions. Twenty mg fresh weight lots of L. longiflorum were sown in sterile Petri dishes containing 20-mL aliquots of sterile DICKINSON's (1965) medium without tetracycline. Prior to the addition of medium, AFB $_2$  or AFG $_1$  (Applied Sciences Lab, Inc.) was added to the Petri dishes to yield 15 µg/mL stocks of each toxin. The stocks of 10 mL or less were diluted to 20 mL with DICKINSON's medium to yield toxin concentrations of 7.500, 3.750, 1.875, 0.938, 0.468 and 0.234 µg/mL. To obtain AFB $_1$  (Calbiochem) -media concentrations of 5, 10, 15, 20 and 25 µg/mL, DICKINSON's media containing 30 µg/mL toxin and with or without 3.0 mM KH $_2$ PO $_4$  were diluted. The toxin which was dissolved in acetone was added to the media which were then autoclaved to remove the solvent. Pollen was germinated at 24 + 2°C for 2, 4 and 8 h except for pollen sown in media containing either AFB $_1$  or AFB $_2$  which were germinated for 4 h.

At each of these times, media aliquots containing germinated pollen were removed from the dishes to shell vials each of which had been prefilled with 100 mL formaldehyde. Then, the vials were maintained at  $4^{\circ}\text{C}$  until pollen germination and tube lengths could be quantified.

Quantification of % germination and tube elongation. Drops were removed from each vial and the % germination and tube lengths were determined with a microscope equipped with an ocular micrometer (JONES et al. 1980; DASHEK et al. 1981).

Chromatography of aflatoxins. AFB1, AFB2 and AFG1 purities were assessed by chromatography on 250- $\mu$ m thick Adsorbosil-1 thin layer chromatography (TLC) plates (Applied Sciences Lab, Inc.). Plates were developed in 5% chloroform in methanol and then viewed with a UV source to visualize the aflatoxins. To verify media AFB1, AFB2 and AFG1 stock concentrations, fluorescent spots on TLC plates were compared densitometrically to reference samples which were spotted simultaneously with the media samples.

## RESULTS AND DISCUSSION

Pollen germination and tube length bioassays of AFB 1. The dose-response curves relating log percent germination and tube length to AFB 1-medium concentration are presented in Fig. 2A and B, respectively. When  $\mathrm{KH_2PO_4}$  was withheld from the medium, the log percent germination remained nearly constant from 5-30  $\mu\mathrm{g/mL}$  (Fig. 2A). In contrast, when  $\mathrm{KH_2PO_4}$  was included within the medium, the log percent germination did not markedly change until the AFB 1-medium concentration was raised to 20  $\mu\mathrm{g/mL}$ . A 20 % decrease in log percent germination occurred when the AFB 1-medium concentration was changed from 20-30  $\mu\mathrm{g/mL}$ . Linear regression analysis yielded a correlation coefficient of 0.98.

In the absence of  $\mathrm{KH_2PO_4}$ , log tube length became less

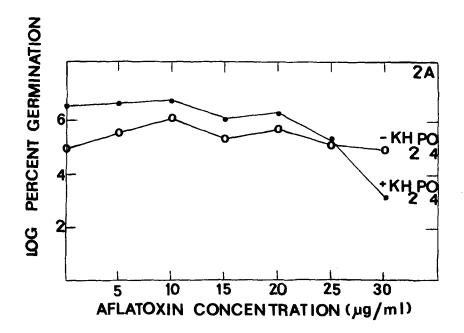


Fig. 2A. Dose-response curve relating log percent germination to AFB<sub>1</sub>-medium concentrations.

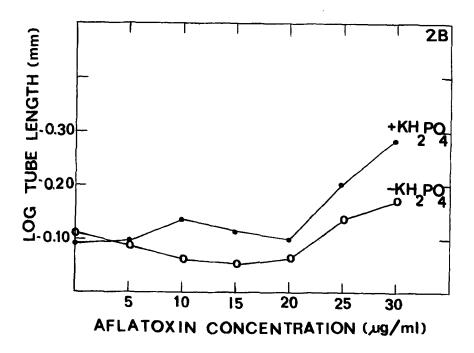


Fig. 2B. Dose-response curve relating log tube length to  $\mbox{\rm AFB}_1\mbox{-medium}$  concentrations.

negative (Fig. 2B) as the concentration was raised to 2  $\mu$ g/mL but more negative as the AFB<sub>1</sub> concentration was elevated from 20-30  $\mu$ g/mL. A greater AFB<sub>1</sub>-induced inhibition of log tube length occurred when tubes were cultured in medium containing KH<sub>2</sub>PO<sub>4</sub> than without KH<sub>2</sub>PO<sub>4</sub>. Correlation coefficients as determined by regression analysis of AFB<sub>1</sub>-promoted inhibitions of log tube length between 20-30  $\mu$ g/mL were 0.99 and 0.98, respectively, for pollen germinated in medium containing and lacking KH<sub>2</sub>PO<sub>4</sub>. The dose-response curves presented within Fig. 2A relating log percent germination to aflatoxin-medium concentration demonstrate that log percent germination could possibly be developed as an AFB<sub>1</sub> bioassay between 20 and 30  $\mu$ g/mL provided that 3.0 mM KH<sub>2</sub>PO<sub>4</sub> is included within the medium. However, it seems that a greater inhibition of tube elongation occurred when the medium was provided with KH<sub>2</sub>PO<sub>4</sub>.

Pollen germination and tube length bioassays of AFG . The log percent germinations at 0.468, 0.937, 1.875, 3.750, 7.500 and 15.000 µg/mL were 1.74, 1.61, 1.66, 1.19, 1.07 and 1.15 times greater than that of the control for 2 h germinated pollen (Fig. 3A). A correlation coefficient of 0.92 was obtained when a linear regression analysis was performed upon the decrease in log percent germination which occurred between AFG medium concentrations of 0.468 and 7.500 µg/mL.

A 9% increase, 8% decrease, 42% diminution, 20% decline, 34% reduction and 7% enhancement in log tube length from that of the control were observed at medium concentrations of 0.468, 0.937, 1.875, 3.750, 7.500 and 15.000  $\mu$ g/mL, respectively (Fig. 3B).

At 4 h of culture, the log percent germination increased 1.20 times over that of the control as the AFG<sub>1</sub>-medium concentration was raised to 1.875 µg/mL (Fig. 3A). The enhancement was 1.14 when the concentration was elevated to 3.750 µg/mL. At 7.500 µg/mL the log percent germination approximated that of the control, but at 15.000 µg/mL it was 6% less. Linear regression analysis of the decrease in log percent germination which occurred between 1.875 and 15.000 µg/mL resulted in a correlation coefficient of 0.94.

Whereas a 75% decrease in log tube length was seen at 7.500  $\mu$ g/mL, supplying the germination medium with 0.468, 0.937, 1.875, 3.750 and 15.000  $\mu$ g/mL promoted tube elongation 2.05, 1.25, 1.07, 1.01 and 1.29 folds, respectively, during a 4 h culture (Fig. 3B).

The log percent germination was 1.18 times greater than that of the control at 0.468  $\mu g/mL$  but declined to nearly that of the control upon raising the AFG1-medium concentration to 3.750  $\mu g/mL$  during an 8 h incubation (Fig. 3A). The log percent germinations were 8 and 15 % less than that of the control at 7.500 and 15.000  $\mu g/mL$ . A correlation coefficient of 0.93 was obtained when a linear regression analysis was performed for the decline which resulted between 0.468 and 15.000  $\mu g/mL$ .

A dose-response curve with a clearly defined trend for

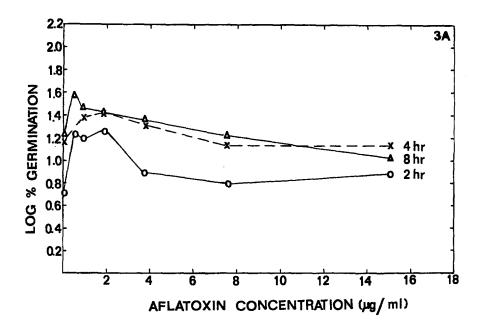


Fig. 3A. Dose-response curve relating log percent germination to AFG<sub>1</sub>-medium concentrations.

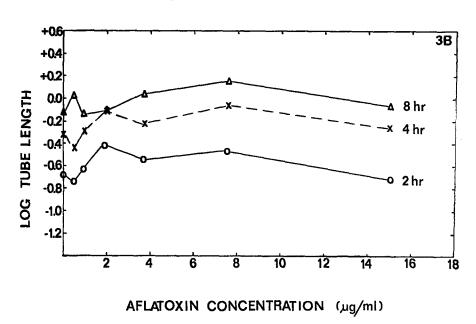


Fig. 3B. Dose-response curve relating log tube elongation to  ${\tt AFG}_1\text{--medium}$  concentrations.

concentration-dependent induced effects on log tube elongation was not observed for pollen which was germinated for 8 h. This together with unsatisfactory correlation coefficients (<0.9) preclude the possibility of developing an AFG<sub>1</sub> bioassay which is based upon tube elongation. Although inclusion of 0.468-15.000  $\mu g/mL$  AFG<sub>1</sub> within the culture medium did not inhibit log percent germination at 2, 4 or 8 h, the linear (r = 0.9) declines (Fig. 3A) in log percent germination from a maximum stimulated level (0.468  $\mu g/mL$  - 2 and 8 h; 1.875  $\mu g/mL$  - 4 h) could possibly be developed as bioassays for the detection and quantification of AFG<sub>1</sub>.

Pollen germination and tube length bioassays of AFB<sub>2</sub>. Fig. 4A demonstrates that the log percent germination remained nearly constant as the medium AFB<sub>2</sub> concentration was enhanced during a 4 h culture precluding the employment of log percent germination as a bioassay for AFB<sub>2</sub>, at least over the concentration range of 0.938 – 15.000 µg/mL. In contrast, log tube length exhibited a rise as the medium concentration was brought to 0.938 µg/mL and then decreased from the stimulated level to a level which approximated that of control at 15.000 µg/mL (Fig. 4B). Linear regression analysis of the diminution from the stimulated level yielded a correlation coefficient of <0.90.

Comparison of pollen germination and tube elongation responses to AFB<sub>1</sub>, AFB<sub>2</sub>, and AFG<sub>2</sub>. When the data within Fig. 2A, 3A and 4A are compared, it is apparent that none of the toxins inhibited germination on a log percent germination basis at  $^{<15}$  µg/mL. However, as little as 0.468 µg/mL AFG<sub>1</sub> appeared to stimulate germination. This concentration is considerably less than those which affect the germination of most plant systems (Table 1). The only systems whose germinations appear to be affected by low toxin concentration are Glycine max, cv. 'Essex' seeds (JONES et al. 1980) and Onoclea sensibilis spores (CAHILL et al. 1978) and in those systems the toxin which was tested was AFB<sub>1</sub> rather than AFG<sub>1</sub>.

Comparison of the data within Fig. 2B, 3B and 4B reveals that none of the toxins inhibited log tube length at <15  $\mu g/mL$ . As in the case of germination, AFG $_1$  concentrations of <2  $\mu g/mL$  stimulated tube elongation. Such stimulation also occurred with AFB $_2$ . To our knowledge, such low toxin concentrations have not been reported to affect the growth of those plant systems tested thus far (Table 2) for possible aflatoxin-induced alterations of growth and development.

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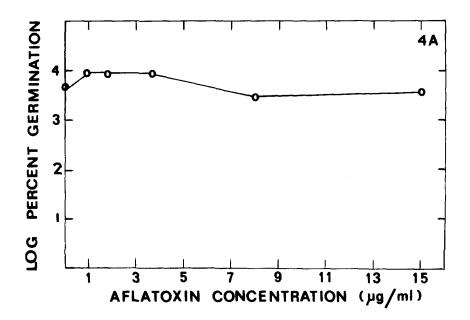


Fig. 4A. Dose-response curve relating log percent germination to  ${\rm AFB}_2{\rm -medium}$  concentration.

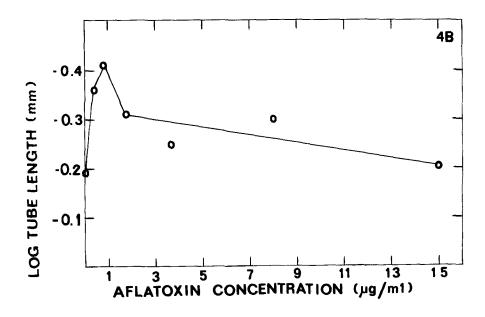


Fig. 4B. Dose-response curve relating log tube elongation to  ${\rm AFB}_2{\rm -medium}$  concentration.

Table 1. Comparison of Germination of Various Seeds Exposed to Aflatoxin

Seed	Response	Aflatoxin Concentration	Investigator(s)
Arachis hypogaea	No significant effect	31.50 µg/mL mixed aflatoxins for 72, 144, 168 and 240 h	DASHEK et al. (1981)
Avena sativa	No significant effect for cvs. 'Norline', 'Wind-sor' or 'Moregrain'; 20% inhibition for cvs. 'Coker' and 'Roanoke'	31.50 µg/mL mixed aflatoxins for 65, 89 and 117 h	DASHEK et al. (1981)
Cuminum cyminum	5% inhibiton after 8 days of imbibition	2.50 and 5.00 $\mu \mathrm{g/mL}$ AFB $_{\mathrm{l}}$	LLEWELLYN et al. (submitted)
Glycine max, cv. 'Essex'	% inhibitions were 5, 20, 40 and 80 or 6, 4, 13, 19 for listed concentrations at 18 and 36 h, respectively	0.38, 2.90, 5.80 and 11.60 μg/mL AFB <sub>1</sub> for 18 and 36 h	JONES et al. (1980)
Hordeum vulgare	No significant effect at 39, 63 and 89 h for cvs. 'Barsoy', 'Volbar' and 'Henry';	significant effect 31.50 µg/mL mixed aflatoxins 39, 63 and 89 h for 39, 63, and 89 h cvs. 'Barsoy', class', and 'Henry';	DASHEK et al. (1981)

Comparison of Germination of Various Seeds Exposed to Aflatoxin Cont'd. Table 1.

Seed	Response	Aflatoxin Concentration	Investigator(s)
	17-23% inhibitions for 'Surry' and 'McNair' at 39, 63 and 89 h, respectively		
Lactuca sativa	No inhibition at 1,000 µg/mL in one cv. nor by 100 µg/mL in 29 other cvs.	100-1000 $\mu \mathrm{g/mL}$ AFB $_1$	CRISAN (1973a)
Lepidium sativum	No impairment;35, 90 and 100% inhibitions, respectively	1,2,5, and 10 µg/mL aflatoxins 25,50 and 100 µg/mL aflatoxins	SCHOENTAL & WHITE (1965)
Lilium longiflorum,	No inhibitory effect; 10.6, 27.3 and 45.1% inhibitions	5-30 $\mu g/mL$ AFB $_1$ without 3.0 mM KH $_2$ PO $_4$ 15, 25 and 30 $\mu g/mL$ AFB $_1$ with KH $_2$ PO $_4$	JONES et al. (1980) DASHEK et al. (1981)
Onoclea sensibilis Pimpinella anisum	6.7, 7.8, 27.0, 32.6 and 43.8% inhibitions, respectively <5 % inhibition after 8 days of imbibition	0.78, 1.56, 2.34, 3.13 and 3.90 µg/mL 2.50 µg/mL AFB <sub>1</sub>	CAHILL et al. (1978) LLEWELLYN et al. (Submitted)

Aflatoxin Cont'd.	Investigator(s)	CRISAN (1973b)		JACQUET et al. (1971)	ADEKUNLE & BASSIR (1973)	DASHEK et al. (1981)		
Comparison of Germination of Various Seeds Exposed to Aflatoxin Cont'd.	Aflatoxin Concentration	500 $\mu_{\rm S}/{\rm mL}~{ m AFB}_{ m J}$ 100 $\mu_{\rm S}/{\rm mL}~{ m AFB}_{ m J}$		20 μg/mL mixed aflatoxins	50 μg/mL	0.35, 0.73, 1.50 and 2.00	5.80 and 11.60 µg/mL mixed aflatoxins	
)	Response	no inhibition no effect		Variable % inhibition (as high as 100% in Phalaris canariensis)	100% inhibiton	No effect	23 and 25% reductions	
Table 1.	Seed	Species of Cruci-	(19 plants belong- ing to 11 species)	Variety of seeds	Vigna sinensis	Zea mays		

Comparison of Growth Responses of Various Plant Parts to Aflatoxins Table 2.

Investigator(s)	DASHEK et al. (1981)	DASHEK et al. (1981)	REISS (1969)	SULLIVAN & IKAWA (1972)	LLEWELLYN et al. (Submitted)	
Aflatoxin Concentration	31.50 µg/mL mixed aflatoxins for 72, 144, 168 and 240 h	31.50 µg/mL mixed aflatoxins for 65, 89 and 117 h	100 and 300 ppm	30 ppm $1~\mu \mathrm{g/mL~AFB}_{\mathrm{l}}$	$2.50~\mathrm{ug/mL~AFB_1}$ for 2, 4 8 days	5.00 $\mu g/mL$ AFB for 2, 4 8 days
Response	No significant effect	% inhibition of root elongation ranged from 4.3 to 68.8%, respectively	death and prevention of growth of upper leaves and floral buds	similar to nontreated inhibitions of 4 strains	% difference from control was 0, 13 and 13, respectively	% difference from control was 0, 0 and 56, respectively
Plant	Arachis hypogaea	Avena sativa	Carralluma freri	Chlorella pyrenoidosa	Cuminum cyminum	

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Table 2.

Plant	Response	Aflatoxin Concentration	Investigator(s)
Glycine max, cv. 'Essex'	Non-treated, excised roots followed a sigmoidal growth curve with a dry wt. increase from 100% (0 h) to 108.5% (24 h); 4.5% dry wt. decline at 4 h, increase to 101.5% (8 h), and decrease to 99% (12 h)	$_{ m 20}$ µg/mL $_{ m AFB}_{ m 1}$	YOUNG et al. (1978)
Glycine max, cv.	% inhibition of attached root elongation was 14 (48 h) and 26 (140 h)	$2.90~\mu \mathrm{g/mL~A^FB}_{\mathrm{J}}$	JONES et al. (1980)
	% inhibition of attached root elongation was 21 (48 h) and 25 (140 h)	5.80 µg/mL AFB]	
	% inhibition of attached root elongation was 36 (48 h) and 50 (140 h)	11.60 $\mu \mathrm{g/mL}$ AFB $_\mathrm{l}$	
Hordeum vulgare	% inhibitions of root elongation ranged from 22.4 to 62.2	31.50 µg/mL mixed aflatoxins for 39 and 89 h	DASHEK et al. (1981)

Comparison of Growth Responses of Various Plant Parts to Aflatoxins. Cont'd. Table 2.

or(s)	7)	1)		1. (1980)	JACQUET et al. (1971)	LLEWELLYN et al. (Submitted)	
Investigator(s)	REISS (1977)	REISS (1971)		JONES et al. (1980)	JACQUET et	LLEWELLYN	
Aflatoxin Concentration	100 $\mu g/mL$ AFB $_1$	1 $\mu g/mL$ AFB <sub>1</sub>	10 and 100 $\mu\mathrm{g/mL}$ AFB $_1$	25 and 30 $\mathrm{kg/mL}$ AFB with 3.0 mM $\mathrm{KH_2PO_4}$	50 µg/mL mixed aflatoxins	2.50 $\mu g/mL$ AFB <sub>1</sub> for 2, 4 and 8 days	5.00 $\mu g/mL$ AFB <sub>1</sub> for 2, 4 and 8 days
Response	Inhibition of root elongation by approximately 50%	No inhibition of hypocotyl elongation No inhibition of radicle elongation	14.2 and 58.0% inhibtion of hypocotyl elongation, 23.9% inhibitions of radicle elongation	23 and 36% inhibitons of pollen tube elongation, respectively	Radicle elongation impaired by 50%	% difference from control was 10, 83 and 71, respectively	% difference from control was 10, 82 and 82, respectively
Plant	Kalanchoe diagrem- ontia	Lepidium sativum	750	Lilium longiflorum	Phalaris canariensis	Pimpinella anisum	

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